# PBMC Immunophenotyping (Panels 1-5)

PART 1: Staining Protocol (Panels 1-4)

#### **REAGENTS AND BUFFER**

- 1. 1x DPBS (Invitrogen, Catalogue number 14190)
- 2. FACS buffer (1X PBS + 2% FCS + 2mM EDTA)
- 3. BD Cell fix (BD Biosciences, Catalogue number 340181)
- 4. BD Brilliant violet stain buffer (BD Biosciences, 566385)
- 5. Live dead dye (BD Biosciences, Catalogue number 565388)

## **MATERIALS**

- 1. P10, P20, P200 and P1000 filter tips
- 2. Beaker for waste

#### **EQUIPMENT**

- 1. BSL3 facility
- 2. BSC2 safety cabinet
- 3. Centrifuge
- 4. P10, P20, P200 and P1000 pipettes
- 5. Multichannel pipette
- 6. Dispensing troughs for multichannel pipetting
- 7. BD LSR Fortessa X20

## Inside BSL3

Samples are in 96 well V-bottom plates as detailed in 'Sample preparation protocol'.

Antibody panels can be found in the 'Set up and panels' document.

- 1. Centrifuge cells for 5 minutes at 400g at RT.
- 2. Prepare Live dead (L/D) dye to working concentration (1:4000 dilution of stock prepared according to manufacturer's instructions in 1x PBS)
- 3. Remove supernatant, and gently resuspend the cell pellets in 100  $\mu$ l L/D working solution by pipetting up and down. Incubate for 20 minutes at RT, protected from light.
- 4. After the L/D incubation step is over, centrifuge cells for 5 minutes at 400g at RT.
- 5. Remove supernatant from cells, and gently resuspend the cell pellet in 200μl FACS buffer by pipetting up and down.
- 6. Centrifuge cells for 5 minutes at 400g at RT.
- 7. Remove supernatant from cells, and gently resuspend the cell pellet directly into 100  $\mu$ l of the antibody master mix prepared in BD Brilliant Violet Stain Buffer and incubate for 30 minutes at RT

- 8. Add 100  $\mu$ l of FACS buffer to wells and centrifuge cells for 5 minutes at 400g at RT.
- 9. Remove supernatant from cells, and gently resuspend the cell pellet in  $200\mu l$  of FACS buffer by pipetting up and down.
- 10. Centrifuge cells for 5 minutes at 400g at RT.
- 11. Fixation: Remove supernatant from cells, and gently resuspend the cell pellet in 100μl of FACS buffer. Add 100μl of fixation buffer and incubate cells for 1 hour protected from light.
- 12. Centrifuge cells for 5 minutes at 400g at RT.
- 13. Remove supernatant from cells, and gently resuspend the cell pellet in  $200\mu l$  of FACS buffer by pipetting up and down.
- 14. Centrifuge cells for 5 minutes at 400g at RT.
- 15. Remove supernatant from cells, and gently resuspend the cell pellet in  $200\mu l$  of FACS buffer by pipetting up and down.
- 16. Store at 4°C and acquire the next day on BD LSR Fortessa X20